Production of Probiotic Nutritive Beverages Fortified with Bioactive Compounds and Antioxidants of Pumpkin and Strawberry Pulps

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IN THIS STUDY, probiotic nutritive beverages were formulated by mixing fermented milk permeate with pumpkin or strawberry pulps at a ratio of (1:1), and stored at 4±1°C for 30 days. Milk permeate was fermented with the use of a 2% mixed starter culture (1:1:1) containing Lactobacillus delbrueckii subsp bulgaricus, an exopolysccharide (EPS) strain of Streptococcus thermophilus, and a probiotic organism Bifidobacterium longum. Changes in antioxidant activity and bioactive compounds as well as the chemical, rheological, microbiological and organoleptical characteristics of the resulting functional beverages were studied. The total solid, ash, fat, protein, fiber and acidity contents were slightly increased, while the total carbohydrate, antioxidant activity, total phenols, total flavonoids, ascorbic acid, anthocyanin and carotenoids contents and pH value were decreased during cold storage. The addition of pumpkin and strawberry pulps greatly improved but variably the Ca, P, Na, K, Mg, Fe, Cu and Zn contents in the prepared beverage. Lactic acid bacteria (LAB), Str. thermophilus, Lb. delbrueckii subsp. bulgaricus or Bif. longum counts were not detected in prepared beverages from heated fermented permeate during cold storage, while the total viable bacteria, LAB, Str. thermophilus, Lb. delbrueckii subsp. bulgaricus and Bif. longum counts gradually decreased in beverages produced from unheated fermented milk permeate during cold storage.

Keyword: Functional beverages, Milk permeate, Pumpkin, Strawberry, Probibtic bacteria, Exopolysccharide, Antioxidant activity, Bioactive compounds.

A fermented dairy beverage was made with milk and whey, and characterized as an important source due to the presence of protein with a high biological value that is mostly derived from whey, a major raw material (Sanmartin *et al.*, 2011). The production of dairy beverages has been increasing worldwide due to their simple production technology and wide acceptance by consumers; they have been characterized as an alternative use for the whey from cow milk resulting from cheese production (Hernandez-Ledesma *et al.*, 2011). In addition, lactic acid bacteria (LAB) produce different inhibitory substances that can prolong the shelf life of the fermented products. Reduction of lactose content in permeate by LAB fermentation is an effective way to avoid lactose intolerance; a serious problem for a significant sector of consumers (Geilman *et al.*, 1992).

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Probiotics are alive microorganisms which when administered in adequate amounts confer a health benefit on the host (FAO/WHO, 2002) by improving microbial balance in the host's gut flora and defenses against pathogenic microorganisms. Other benefits attributed to probiotics include prevention of cancer, stimulation of the immune system, lowering of serum cholesterol levels, and improvement of vitamin synthesis (Heenan *et al.*, 2004). The species which are most frequently used as probiotics belong to the genera *Lactobacillus* and *Bifidobacterium* (Isolauri, 2004).

Food produced from plants (fruits and vegetables) abounds with natural biologically active compounds, such as polyphenols, vitamin C or β -carotene, anthocyanins, flavones, have their antioxidant properties, are of great value to human health. It is thought that an adequate level of antioxidants supplied with one's diet induces immunological processes and increases defensive abilities of cells in proper way (Kalt, 2005). A well-balanced diet that will strengthen the immune system is now of great significance.

The photochemical antioxidants such as carotenoids, flavonoids and phenols ... etc., have potential health roles in the reduction of platelet aggregation, blood pressure, cardiovascular of disease and a role in modulation of cholesterol synthesis and absorption (Li, 2008). Moreover, fruits and vegetables contain protein bound polysaccharides which may increase the levels of serum insulin, reduce the blood glucose levels and improve tolerance of glucose and hence could be developed as new anti-diabetic agents (Li *et al.*, 2005).

Pumpkin (*Cucurbita maxima*) is an excellent source of carotenoids, ascorbic acid, polysaccharides, mineral compounds (K, Ca, Mg and Fe), starch (1.5-20%), and pectin (4.8-12.8%). Pumpkin contains significant amounts of beta and alpha carotene and provides a very good amount of beta cryptoxanthin (Li, 2008).

Strawberries (*Fragaria xananassa*) are rich in phenols and particularly a class of phenols called anthocyanins. The anthocyanins in strawberries give the fruit its red colour and acts as a powerful antioxidant that protects the bodies' cells from free radical damage. Strawberry nutrition is an excellent source of vitamin C and B3, carbohydrates, organic acid, flavonoids, pectin and mineral compounds (Cordenunsi *et al.*, 2002 and Li, 2008).

The predominant types of whey or permeate drinks are based on blends of fruit juices and unprocessed or modified whey. Today beverages are one of the most popular vehicles to deliver nutrients and phyto-nutrients for health benefits. Addition of vitamins and micronutrients to food stuffs is practiced in a number of countries' to combat deficiency diseases (Wrick, 2003).

The objective of this research study consisted of determining the impact of strawberry and pumpkin pulps on the nutritional value, bioactive compounds; antioxidant activity and sensory evaluation of fermented milk permeate beverages with probiotic bacteria.

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Materials and Methods

Materials

Milk permeate was obtained from the ultrafiltration of buffalos milk using Carbo-sep, UF unit (SFEC, France) at the Animal Production Research Institute, Agriculture Research Center, Egypt. Commercial grade of granulated sugarcane was obtained from the local market, Egypt.

Pumpkin (*Cucurbita maxima* L) was obtained from the farm of Fac. of Agric. Moshtohor, and strawberry (*Fragaria xananassa*) was purchased from the local market, Egypt.

Pure cultures of *Lactobacillius delbrueckii* subsp *bulgaricus* DSM 20080, *Bifidobacterium longum* DSM 20088 and exopolysaccharides (EPS) producing strain *Streptococcus thermophilus* ASCC 1275 were kindly obtained from the Institute of Microbiology, Federal Research Center for Nutrition and Food, Kiel, Germany.

Methods

Preparation of fruit pulp

Pumpkin and strawberry were washed thoroughly with tap water. Pumpkin was hand peeled, seeds were carefully removed and the flesh was cut into "small pieces". The prepared fruits flesh was then blanched at 80°C for 10 min, according to Gupta (1998).

Fermentation of milk permeate

Milk permeate was warmed up to 40° C, sucrose was added at the ratio of 5g/100 ml, heated to 85°C for 15 min in a water bath and then rapidly cooled. The sweetened milk permeate was inoculated with 2% of mixed starter culture (1:1:1): *Lb. delbrueckii* subsp *bulgaricus*, *Bif. longum* and *Str. thermophilus*, incubated at 42°C until the pH was decreased to 5 and rapidly cooled to $4\pm1^{\circ}$ C.

Preparation of the functional beverages

Preliminary experiments were carried out to select the best formulation for the target beverages when fermented permeate (25, 50 and 75%), was mixed with pumpkin and strawberry pulps (75, 50 and 25%), respectively and sensory evaluated. Results showed that beverage formulations containing equal quantities of the fermented permeate and pumpkin and strawberry pulps yielded the highest sensory scores, which were chosen for further study.

Fermented milk permeate was divided into three portions. Functional beverages were prepared as follows:

- Firstly (4 kg) of unheated (C1) and heated (C2) fermented permeates $(85^{\circ}C/15min)$ were kept as controls.

- Secondly (2 kg) of unheated (T1) and heated (T3) fermented permeates (85°C/15min) were mixed with an equal volume of pumpkin pulp.

- Thirdly (2 kg) of unheated (T2) and heated (T4) fermented permeates $(85^{\circ}C/15min)$ were mixed with an equal volume of strawberry pulp.

The prepared beverages were filled into sterilized bottles, stored at 4 ± 1 °C and were analyzed chemically, microbiologically and were also sensory evaluated when fresh and after 10, 20 and 30 days of cold storage. Also, the apparent viscosity of the fresh beverages was measured. The experiment was repeated three times and all analyses were carried out in duplicate.

Chemical analysis

The total solids, fat, ash, total protein, titratable acidity, pH and ascorbic acid contents were determined according to the methods described by AOAC (2000). The total sugar contents were determined by Shaffer and Hartman method as described in the AOAC (2000). Crude fiber was determined by Weende method (AOAC, 2000). Carotenoids were determined according to Harvey and Catherine (1982). Minerals contents were determined according to AOAC (2000) using Perkin-elmer, 2380 Atomic absorption spectrophotometry. Antioxidant activity, total phenolic, flavonoids and anthocyanin contents were determined according to the methods described by Prieto *et al.* (1999), Shiri *et al.* (2011), Bor *et al.* (2006) and Cordenunsi *et al.* (2002), respectively. The amino acids profile of the functional beverage was performed following the protocol of Walsh and Brown (2000).

Microbiological examinations

Total lactic acid bacterial (LAB), yeasts and moulds; the total viable bacterial and colifrom bacterial counts were determined according to Elliker *et al.* (1956), IDF (1990), IDF (1991) and APHA (2001), respectively. *Lb. delbrueckii* subsp. *bulgaricus, Str. thermophilus* and *Bifidobacterium* sp. were enumerated according to the methods described by Ryan *et al.* (1996) and Martin & Chou (1992), respectively.

Sensory evaluation

The sensory evaluation of beverages was done by a taste panel of 10 experienced panelists from the staff-members of Food Science Department, Faculty of Agriculture, Moshtoher, Benha Univ. The samples were evaluated for taste, odour, body and texture, appearance, sweetness, acidity and colour out of 20, 20, 20, 10, 10, 10 and 10 score points (Fellers *et al.*, 1986 and Bodyfelt *et al.*, 1988).

Measurement of viscosity

The apparent viscosity of the prepared beverages was measured using a Brookfield viscometer Model DV11 + Pro (Brookfield unit, MA, USA) at 25°C with a rotation speed of 60 rpm. The results are presented in milli-pascal seconds (mPa.s).

Statistical analysis

Statistical analysis for the obtained data was performed according to the methods of Clarke and Kempson (1997).

Results and Discussions

Composition of used ingredients

Chemical composition, antioxidant activity and bioactive compounds of raw materials used for the manufacture of the prepared functional beverages are presented in Table 1. Pumpkin pulp recorded the highest (P≤0.05) value of total solids, ash, total sugars, fiber and carotenoids contents while, milk permeate showed the lowest. In addition, milk permeate yielded the highest (P≤0.05) pH value. Fresh strawberry pulp contained generally high fat, protein, titratable acidity, antioxidant activity, total phenols, total flavonoids, ascorbic acid and anthocyanin contents (P≤0.05). Total phenols, total flavonoids, ascorbic acid and anthocyanin contents were not detected in milk permeate. The results of chemical composition for ingredients used for the preparation of produced functional beverages were in agreement with those obtained by Nawirska-Olszanska et al. (2011) and Abou El Samh et al. (2013).

Components	Pumpkin pulp	Strawberry pulp	Milk permeate
Total solids %	10.96 ^a	9.89 ^b	5.68 ^c
Ash %	0.47 ^a	0.45 ^a	0.26 ^b
Fat %	0.18 ^b	0.25^{a}	0.10 ^c
Protein %	0.59 ^b	0.71 ^a	0.22 ^c
Total sugars %	8.06 ^a	7.12 ^b	5.01 ^c
Fiber %	2.00^{a}	1.69 ^b	ND
Titratable acidity % as lactic acid	0.21 ^b	0.70^{a}	0.09 ^c
pH values	6.09 ^b	3.75 [°]	6.44 ^a
Antioxidant activity (mg 100g ⁻¹)	80.81 ^b	254.20 ^a	10.95 ^c
Total phenols (mg 100g ⁻¹)	42.94 ^b	200.15 ^a	ND
Total flavonoids (mg 100g ⁻¹)	3.12 ^b	16.23 ^a	ND
Ascorbic acid (mg 100g ⁻¹)	12.32 ^b	85.45 ^a	ND
Anthocyanin (mg 100g ⁻¹)	5.45 ^b	55.34 ^a	ND
Carotenoids (mg 100g ⁻¹)	10.54 ^a	1.14 ^b	ND

TABLE 1. Chemical composition, antioxidant activity and bioactive com	mpounds of
raw materials used in prepared functional beverages.	

ND = Not detected

^{a-c} Different letters in the same row indicate significant statistical differences (Duncan's test $P \le 0.05$)

Physicochemical composition of prepared functional beverages

Table 2 summarizes the physicochemical composition of the prepared functional beverages during storage up to 30 days at 4±1°C. The total solids and ash contents increased slightly in all treatments during the storage periods. Generally, there were significant differences (P≤0.05) between total solids and ash contents of beverage treatments during the storage. The slight increase of total solids during storage may be attributed to the loss of some moisture content during the cold storage. These results are in accordance with those of Hashmi et al. (2011) and Atallah (2015).

Treatments	Total solids%	Ash %	Fat %	Protein %	Total carbohydrates %	Acidity %	рН	Fiber %
Fresh								
C1	9.09 ^{fA}	0.27 ^{cC}	0.10 ^{bA}	0.37 ^{cB}	7.59 ^{fA}	0.85 ^{bC}	4.17 ^{dA}	ND
C2	9.15 ^{eA}	0.28 ^{cC}	0.12 ^{aA}	0.39 ^{bB}	7.68 ^{eA}	0.75 ^{eC}	4.49 ^{bA}	ND
T1	10.09 ^{bA}	0.33 ^{bC}	0.10 ^{bA}	0.41 ^{bB}	7.95 ^{bA}	0.81 ^{cC}	4.29 ^{cA}	0.60^{dA}
T2	9.97 ^{dA}	0.38 ^{aC}	0.12 ^{aA}	0.50 ^{aB}	7.71 ^{dA}	0.89 ^{aC}	3.82 ^{eA}	0.85 ^{bA}
T3	10.31 ^{aA}	0.37 ^{aC}	0.15 ^{aA}	0.41 ^{bB}	8.21 ^{aA}	0.65 ^{fC}	4.80 ^{aA}	0.64 ^{cA}
T4	10.01 ^{cA}	0.40 ^{aC}	0.15 ^{aA}	0.53 ^{aB}	7.89 ^{cA}	0.76 ^{dC}	3.95 ^{fA}	1.01 ^{aA}
				10 day	s			
C1	9.30 ^{dA}	0.29 ^{cB}	0.10 ^{bA}	0.38 ^{cB}	7.50 ^{eB}	0.90 ^{bB}	4.02 ^{dB}	ND
C2	9.27 ^{dA}	0.31 ^{cB}	0.12 ^{aA}	0.39 ^{bB}	7.61 ^{dB}	0.77 ^{eB}	4.40 ^{bB}	ND
T1	10.14 ^{bA}	0.33 ^{bB}	0.10 ^{bA}	0.41 ^{bB}	7.90 ^{bB}	0.89 ^{cB}	4.15 ^{cB}	0.60 ^{dA}
T2	10.01 ^{cA}	0.38 ^{aB}	0.12 ^{aA}	0.52 ^{aB}	7.60 ^{dB}	0.94 ^{aB}	3.71 ^{fB}	0.86 ^{bA}
T3	10.38 ^{aA}	0.37 ^{abB}	0.15 ^{aA}	0.41 ^{bB}	8.19 ^{aB}	0.72 ^{fB}	4.72 ^{aB}	0.65 ^{cA}
T4	10.11 ^{bA}	0.41 ^{aB}	0.15 ^{aA}	0.55 ^{aB}	7.80 ^{cB}	0.77 ^{dB}	3.90 ^{eB}	1.02 ^{aA}
				20 day	S			
C1	9.31 ^{dA}	0.31 ^{cA}	0.10 ^{bA}	0.39 ^{cA}	7.50° ^C	1.05 ^{bA}	3.88 ^{dC}	ND
C2	9.28 ^{dA}	0.31 ^{cA}	0.13 ^{aA}	0.41 ^{bA}	7.60 ^{dC}	0.83 ^{eA}	4.27 ^{bC}	ND
T1	10.14 ^{bA}	0.35 ^{bA}	0.10 ^{bA}	0.42 ^{bA}	7.78 ^{bC}	0.99 ^{cA}	3.98 ^{cC}	0.62 ^{dA}
T2	10.07 ^{cA}	0.40 ^{aA}	0.14 ^{aA}	0.54 ^{aA}	7.54 ^{cC}	1.08 ^{aA}	3.61 ^{fC}	0.86 ^{bA}
T3	10.40 ^{aA}	0.40 ^{aA}	0.16 ^{aA}	0.43 ^{bA}	8.11 ^{aC}	0.80 ^{fA}	4.60 ^{aC}	0.68 ^{cA}
T4	10.20 ^{bA}	0.42 ^{aA}	0.16 ^{aA}	0.57 ^{aA}	7.76 ^{bC}	0.85 ^{dA}	3.85 ^{eC}	1.03 ^{aA}
30 days								
C1	9.31 ^{dA}	0.31 ^{cA}	0.10 ^{bA}	0.39 ^{cA}	7.45 ^{cD}	1.17 ^{bA}	3.70 ^{dD}	ND
C2	9.28 ^{dA}	0.32 ^{cA}	0.13 ^{aA}	0.42 ^{bA}	7.56 ^{dD}	0.90 ^{eA}	4.15 ^{bD}	ND
T1	10.16 ^{bA}	0.36 ^{bA}	0.10 ^{bA}	0.43 ^{bA}	7.77 ^{bD}	1.08 ^{cA}	3.76 ^{cD}	0.63 ^{dA}
T2	10.11 ^{cA}	0.40 ^{aA}	0.15 ^{aA}	0.55 ^{aA}	7.48 ^{cD}	1.21 ^{aA}	3.56 ^{fD}	0.86 ^{bA}
T3	10.40 ^{aA}	0.41 ^{aA}	0.15 ^{aA}	0.43 ^{bA}	8.05 ^{aD}	0.85 ^{fA}	4.46 ^{aD}	0.68 ^{cA}
T4	10.20 ^{bA}	0.42 ^{aA}	0.16 ^{aA}	0.58 ^{aA}	7.73 ^{bD}	0.91 ^{dA}	3.75 ^{eD}	1.03 ^{aA}
ND= Not detected								

TABLE 2. Physicochemical composition of the prepared functional beverages during storage periods at 4±1°C.

C1 = Control 1 (unheated fermented milk permeate) C2 = Control 2 (heated fermented milk permeate)

T1 = 50% unheated fermented milk permeate + 50% pumpkin pulp

T2 = 50% unheated fermented milk permeate +50% strawberry pulp

T3 = 50% heated fermented milk permeate + 50% pumpkin pulp

T4 = 50% heated fermented milk permeate + 50% strawberry pulp a-for A-D Different letters in the same colum indicate significant statistical differences (Duncan's test P \leq 0.05)

The fat and protein contents of the produced functional beverages slightly increased (P≤0.05) during cold storage. The fat content of fresh products varied from 0.10 to 0.15% and 0.37 to 0.53% for protein, and then increased slightly up to the end of the storage period.

The total carbohydrate of the prepared functional beverages significantly decreased during storage at 4±1°C up to 30 days and there were significant differences (P≤0.05) between all treatments during storage period. This decrease was expectedly due to microbial activity. These results confirmed those reported by Vahedi et al. (2008).

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Generally, data in the same table showed that the prepared functional beverages from unheated fermented permeate showed higher acidity and lower pH (P \leq 0.05) than beverages prepared from heated fermented permeates throughout the storage period. However, in both heated and unheated beverages, the acidity was increased and pH decreased during cold storage. This can be attributed to the presence of microorganism in all samples as apparent from the total viable counts. The presence of starter culture strains, in unheated beverages may explain the higher acidity and lower pH of these beverages compared with heated beverages. These results confirm those obtained by Hegazi *et al.* (2009).

The fiber content of fresh functional beverages of all treatments varied from 0.60 to 1.01%, and was almost unchanged during storage (Table 2). However, there were significant differences (P \leq 0.05) in fiber content between functional beverages and controls. The fiber in all beverages was less than 5% which may exhibit positive effects on nutrient utilization. Gronowska-Senger *et al.* (1980) found that 5% fiber in the diet had a positive effect on carotene utilization whereas 10 to 20% had a negative effect.

Antioxidant activity and bioactive compounds of functional beverages

Fermented functional beverages containing strawberry and pumpkin pulps can be considered an excellent source of antioxidant activity, total phenols, total flavonoids, ascorbic acid, anthocyanins and carotenoids which are important nutrients in the human diet.

Data in Table 3 showed that the antioxidant activity of the functional beverage containing strawberry was higher than of that containing pumpkin and control samples. However, the antioxidant activity was significantly different (P \leq 0.05) between all treatments during storage periods. Also, the antioxidant activity values were slightly but significantly decreased (P \leq 0.05) during cold storage. The results are in agreement with those given by Zhiyong *et al.* (2015).

Also, the total phenols content of functional beverages was the highest (P \leq 0.05) in strawberry followed by that of pumpkin. The total phenols content ranged from 21.54 to 56.32 mg 100g⁻¹ in the fresh functional beverages. During storage up to 30 days, it decreased from 20.42 to 54.33 mg 100g⁻¹.

In addition, the total flavonoid content in both the functional beverages showed a similar trend to that of total phenols content. The total flavonoid content was significantly higher ($P \le 0.05$) in beverage containing strawberry pulp followed by that containing pumpkin pulp. These results may be due to the higher content of total phenols and flavonoids in strawberry than that in pumpkin pulps.

Treatments	Antioxidant activity	Total phenols	Total flavonoids	Ascorbic acid	Anthocyanin	Carotenoids	
Fresh							
C1	11.02 ^{eA}	ND	ND	ND	ND	ND	
C2	11.03 ^{eA}	ND	ND	ND	ND	ND	
T1	26.98 ^{cA}	21.98 ^{cA}	1.70 ^{cA}	5.10 ^{cA}	1.75 ^{cA}	5.56 ^{aA}	
T2	80.56^{aA}	56.32 ^{aA}	8.54 ^{aA}	21.05 ^{aA}	20.21 ^{aA}	0.51 ^{bA}	
T3	24.65 ^{dA}	21.54 ^{dA}	1.65 ^{cA}	4.23 ^{dA}	1.70 ^{cA}	5.53 ^{aA}	
T4	76.21 ^{bA}	55.89 ^{bA}	8.32 ^{bA}	19.87 ^{bA}	19.87 ^{bA}	0.50 ^{bA}	
			10 days				
C1	10.86 ^{eA}	ND	ND	ND	ND	ND	
C2	10.85 ^{eA}	ND	ND	ND	ND	ND	
T1	26.67 ^{cA}	21.65 ^{cA}	1.66 ^{cA}	4.91 ^{cB}	1.70 ^{cA}	5.54 ^{aA}	
T2	80.51 ^{aA}	55.89 ^{aA}	8.45 ^{aA}	20.88^{aB}	20.01 ^{aA}	0.52 ^{bA}	
T3	24.24 ^{dA}	21.20 ^{dA}	1.59 ^{cA}	4.00^{dB}	1.66 ^{cA}	5.48 ^{aA}	
T4	76.05 ^{bA}	55.55 ^{bA}	8.10 ^{bA}	19.04 ^{bB}	19.79 ^{bA}	0.48 ^{bA}	
			20 days				
C1	10.81 ^{eA}	ND	ND	ND	ND	ND	
C2	10.81 ^{eA}	ND	ND	ND	ND	ND	
T1	26.35 ^{cA}	21.27 ^{cA}	1.54 ^{cA}	4.71 ^{cC}	1.65 ^{cB}	5.49 ^{aA}	
T2	80.40 ^{aA}	55.25 ^{aA}	8.22 ^{aA}	20.54 ^{aC}	19.45 ^{aB}	0.47 ^{bA}	
T3	24.13 ^{dA}	21.00 ^{dA}	1.49 ^{cA}	3.91 ^{dC}	1.51 ^{cB}	5.40 ^{aA}	
T4	76.00 ^{bA}	55.12 ^{bA}	8.01 ^{bA}	18.89 ^{bC}	19.06 ^{bB}	0.42 ^{bA}	
30 days							
C1	10.78 ^{eA}	ND	ND	ND	ND	ND	
C2	10.79 ^{eA}	ND	ND	ND	ND	ND	
T1	26.00 ^{cA}	20.42 ^{dB}	1.43 ^{cA}	4.21 ^{cD}	1.54 ^{cC}	5.38 ^{aA}	
T2	80.02 ^{aA}	54.03 ^{bB}	8.12 ^{aA}	19.90 ^{aD}	18.90 ^{aC}	0.40 ^{bA}	
T3	24.01 ^{dA}	20.52 ^{cB}	1.39 ^{cA}	3.12 ^{dD}	1.37 ^{cC}	5.35 ^{aA}	
T4	75.97 ^{bA}	54.33 ^{aB}	7.95 ^{bA}	18.00 ^{bD}	18.50 ^{bC}	0.37 ^{bA}	
See footnote Table 2.							

TABLE 3. Antioxidant activity and bioactive compounds of prepared functional beverages during storage periods at 4±1°C (mg 100g⁻¹).

^{*}See footnote Table 2. ^{a-e or A-D} Different letters in the same colum indicate significant statistical differences (Duncan's test P≤0.05)

The ascorbic acid contents in functional beverage containing strawberry were higher than that containing pumpkin. However, the ascorbic acid was significantly different (P≤0.05) between all treatments during storage periods. Also, ascorbic acid values were slightly but significantly decreased (P≤0.05) during cold storage.

Also, strawberry functional beverage showed the highest (P≤0.05) anthocyanin contents followed by that containing pumpkin and control samples. The anthocyanins content of the fresh functional beverages ranged from 1.70 to 20.21 mg 100g⁻¹ and decreased gradually during the storage period up to 30 days. The obtained results agree with that of Li (2008).

Data in the same table showed the highest carotenoid contents of beverage functional pumpkin followed by that containing strawberry and control samples. The carotenoids content of the prepared functional beverages decreased gradually (P≤0.05) during storage up to 30 days. Fruits and vegetables have

always been considered an essential part of a healthy diet, with respect to its vitamin C and β -carotene contents (Li, 2008).

Minerals contents of the prepared functional beverages

Table 4 revealed that the highest (P≤0.05) Na, K, Fe, Cu and Mg contents were found in beverages containing strawberry due to the high percentage of these minerals in strawberry pulp. The Ca content was significantly different $(P \le 0.05)$ between all functional beverages, as it ranged between 35.00 to 58.00 mg $100g^{-1}$. While, the P contents (P<0.05) of all functional beverages varied between 19.93 to 29.03 mg 100g⁻¹. Fermented permeate was responsible for the relatively high Ca and P of the prepared beverages due to its high contents of these elements. The trace elements of Mg, Fe, Cu and Zn contents were significantly different (P≤0.05) in the prepared beverages as they ranged from 2.51 to 9.73, 0.41 to 1.02, 0.05 to 0.14 and 0.04 to 0.22 mg 100g⁻¹, respectively, due to the variable contents of these elements in the used ingredients.

From the results of minerals it could be concluded that the prepared functional beverages can be considered a good source for some of these minerals. Trace elements are essential for growth and development, wound healing, immunity and other physiological processes (Miller, 2000). These results closely agreed with those reported by Miller (2000) and Atallah (2015).

Minerals	Fresh functional beverages							
	C1	C2	T1	T2	Т3	T4		
Ca	57.00 ^a	58.00 ^a	35.00 ^c	44.00 ^b	35.41 ^c	43.80 ^b		
Р	28.97 ^a	29.03 ^a	19.93 ^c	24.00 ^b	20.00 ^c	24.60 ^b		
Na	18.59 ^a	18.85 ^a	22.89 ^b	27.00^{a}	23.10 ^b	27.87 ^a		
K	12.41 ^e	13.87 ^d	18.51 ^c	22.50 ^b	18.91 ^c	23.20 ^a		
Mg	2.51 ^f	2.74 ^e	4.42 ^d	9.07 ^b	4.84 ^c	9.73 ^a		
Fe	0.41 ^c	0.43 ^c	0.67 ^b	0.99 ^a	0.69 ^b	1.02 ^a		
Cu	0.05 ^c	0.06 ^c	0.08 ^b	0.10 ^b	0.09 ^b	0.14 ^a		
Zn	0.04 ^d	0.04 ^d	0.19 ^b	0.07 ^c	0.22 ^a	0.08°		
	0.04 ^d							

TABLE 4. Minerals contents of produced functional beverages (mg 100g⁻¹).

*See footnote Table 2. ^{a-f} Different letters in the same row indicate significant statistical differences (Duncan's test P≤0.05)

Amino acid profiles of produced functional beverages

Table 5 shows the results for the concentrations of the essential and non-essential amino acids of fresh functional beverages. Functional beverages containing pumpkin had the highest values of essential and non-essential amino acids compared to that containing strawberry and control samples. Leucine is the major essential amino acid (109.31 mg kg⁻¹) followed by lysine and theronine with mean values 98.24 mg kg⁻¹ and 65.38 mg kg⁻¹, respectively of all functional beverages. In contrary, value for non-essential amino acids of all functional beverages indicated that glutamic acid was present in the highest values (123.03 mg kg⁻¹) followed by aspartic acid (95.89 mg kg⁻¹) and proline (75.64 mg kg⁻¹). The breakdown of leucine takes place in the liver and skeletal muscles (Layman, 2003). It undergoes transamination in the Egypt. J. Food Sci. 43 (2015)

muscles by transferring into glutamine or alanine that ultimately convert to glucose in the liver through gluconeogenesis; a unique pathway for the maintenance of blood glucose level (Borsheim *et al.*, 2003). Hence dietary proteins rich in essential and branched chain amino acids particularly leucine provide health benefits that are not usually observed for diets containing protein from other sources (Wolfe, 2002).

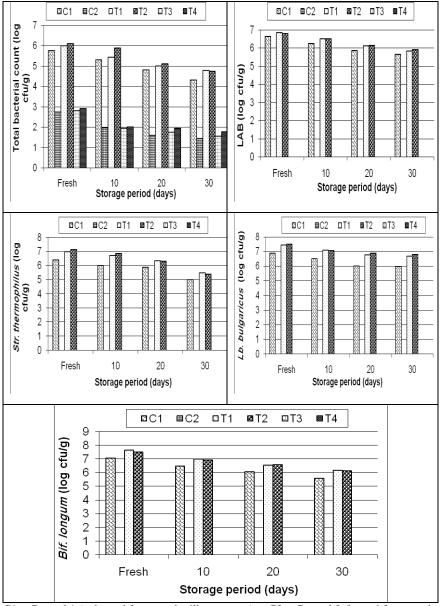
A	Fresh functional beverages						
Amino acids	C1	T1	Т2				
Essential amino acids							
Isoleucine	55.23	65.01	56.32				
Leucine	97.12	109.56	98.88				
Lysine	78.56	98.24	81.84				
Methionine	51.11	65.00	57.98				
Phenylalanine	50.06	65.05	58.34				
Theronine	58.32	67.38	64.30				
Tryptophan	59.94	63.23	60.00				
Valine	51.94	64.44	58.32				
Arginine	54.87	63.21	57.89				
Histidine	40.02	63.98	40.56				
	Non-essential amine	o acids					
Alanine	68.22	73.12	70.00				
Aspartic acid	74.29	95.89	85.32				
Cystein	41.26	46.23	44.39				
Glutamic acid	98.36	123.03	110.23				
Glycine	57.95	72.35	69.65				
Proline	64.29	75.64	70.12				
Serine	54.75	71.23	66.23				
Tyrosine	51.97	60.77	58.77				

TABLE 5. Amino acids	profiles of pro	duced functional	beverages (mg Kg ⁻¹).

*See footnote Table 2.

Microbiological properties

Microbiological examination of the prepared functional beverages for fresh and after 10, 20 and 30 days of storage at $4\pm1^{\circ}$ C for 30 days is shown in Fig 1. There were significant difference (P \leq 0.05) in the microbiological examination between functional beverages treatments and control samples during cold storage. Heat treatment reduced greatly the total viable bacterial count (TVBC) of whey permeate. The TVB and LAB counts gradually decreased up to the end of the storage period. LAB were not detected in heated fermented permeate, and heated fermented beverages throughout the storage period. The *Str. thermophilus* and *Lb. delbrueckii* subsp. *bulgaricus* counts of during storage gradually decreased in all samples. The decrease of counts during storage (P \leq 0.05) may be due to their sensitivity to the developed acidity in the product (Atallah, 2015).



C1 = Control 1 (unheated fermented milk permeate) C2 = Control 2 (heated fermented milk permeate)

T1 = 50% unheated fermented milk permeate + 50% pumpkin pulp

T2 = 50% unheated fermented milk permeate + 50% strawberry pulp

T3 = 50% heated fermented milk permeate + 50% pumpkin pulp

T4 = 50% heated fermented milk permeate + 50% strawberry pulp

Fig .1. Microbiological quality of produced functional beverages during cold storage at $4\pm1^{\circ}C$.

Differences were found in *Bif. longum* counts between beverage samples from different treatments throughout the storage period. *Bif. longum* counts decreased ($P \le 0.05$) at the end of the storage period. The decrease in bifidobacterial counts may be due to the developed acidity during storage periods. These results are in agreement with those obtained by Buriti *et al.* (2014) and Atallah (2015).

The Food and Agriculture Organization/ World Health Organization report (FAO/WHO 2002) which specifics that probiotic foods with health claims must contain per gram at least $10^6 - 10^7$ cfu at the time of consumption to exhibit their potential function.

For fresh functional beverages, coliforms and yeasts & moulds are considered as indicators of plant hygiene, and sanitation quality of such products, and as a warning that the products may constitute a health risk. As a result of high hygiene conditions during processing and storage, coliform bacteria and, yeasts and moulds were not detected in all treatments when fresh and during storage. These results are in agreement with those obtained by Atallah (2015).

Sensory evaluation

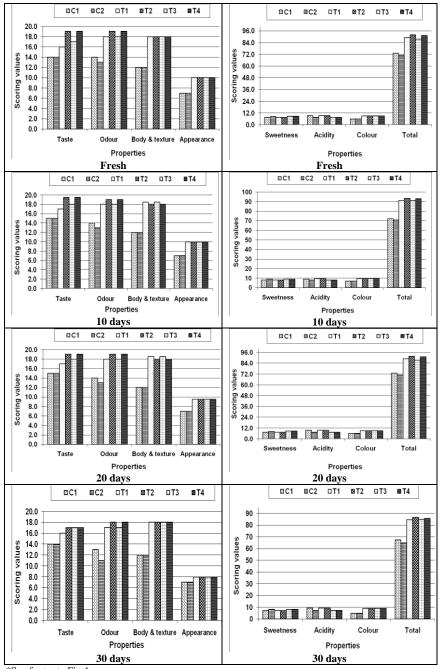
Data in Fig 2 illustrated that all functional beverages containing strawberry and pumpkin have very good scores for acceptability by taste panel compared to control samples when fresh and throughout storage. The functional beverages containing probiotic strains and fruits exhibited improved sensory evaluation due to their high level of produced flavour compounds. Significant differences (P≤0.05) were found in scores for different sensory attributes between all treatments during the storage period. Functional beverage containing strawberry was ranked higher followed by beverage containing pumpkin. It is of interest that beverages based on heated or unheated fermented permeate gained close score points for the different attributes and the total score points. This can be explained on the basis of the slight changes in the composition of the products during storage. During cold storage, the organoleptic scores increased for all treatments after 10 days. No changes were observed among the treatments for all sensory characteristics up to 20 days of storage. After 30 days of cold storage, the same trend was observed for all the tested products with slight decreases in the obtained scores. Similar trends were obtained by Buriti et al. (2014) and Atallah (2015).

Viscosity

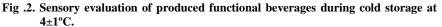
Data in Fig 3 indicated that control samples (C1 and C2) without additives had higher viscosity values than that milk permeate. This can be explained on the use of exopolysaccharide (EPS) producing microorganisms in the fermentation of the permeate. The EPS produced by *Str. thermophilus* strain presumably affects the texture of yoghurt and the functional beverages (Folkenberg *et al.*, 2005).

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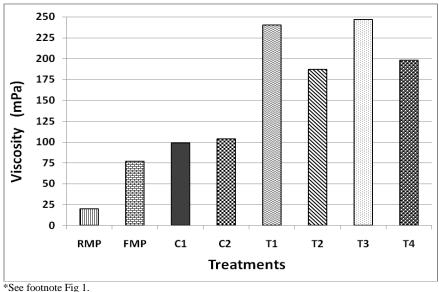
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*See footnote Fig 1.



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RMP= Raw milk permeate FMP= Fermented milk permeate without any additives Fig. 3. Viscosity of raw milk permeate, fermented milk permeate and prepared

functional beverages.

Also, the addition of fruits led to increased viscosity of the prepared beverages compared to control samples. These findings could be related to the higher total solids in functional beverages containing fruit pulp compared to control samples. Also, the addition of pumpkin significantly increased the viscosity of the beverages while strawberry significantly decreased the viscosity of the beverages. The highest viscosity values recorded for pumpkin beverages may be due to the presence of stabilizing agents (dietary fibers) in pumpkin. These have the ability to bind water, which would tend to increase the consistency of the products by increase of the water-binding capacity (Caili et al., 2007). A similar trend was recorded by Atallah (2015).

Conclusion

Pumpkin and strawberry pulps improved the flavour and nutritional value of probiotic beverages. A nutritious functional beverage can be successfully made by mixing fermented milk permeate with pumpkin and strawberry pulp at a ratio of 1:1 with 5% sucrose. The survival of probiotic bacteria during refrigerated conditions for at least 30 days resulted in numbers greater than 10^6 cfu g⁻¹ or 6 log cfu g⁻¹ in some treatments (unheated), which is essential for a product to have probiotic properties. The pulp-enriched samples were characterised by the highest content of antioxidant activity, total phenols, total flavonoids, ascorbic acid, anthocyanins and carotenoids. Functional beverages containing pumpkin had the highest essential and non- essential amino acids concentrations compared

to strawberry and controls. The sensory scores of the functional beverages were high indicating acceptance. Addition of an exopolysaccharide-producing bacterial strain and other materials improved the textural and viscosity properties of the functional beverages. Generally, chemical, rheological, microbiological, organoleptical, antioxidant activity and bioactive compounds indicated that the use of pumpkin and strawberry as flavouring material in functional beverage manufacture is highly recommended. The addition of an EPS producing strain increased the viscosity of the prepared functional beverages.

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References

- Abou El Samh, M.M., Sherein, A.A. and Essam, H.H. (2013) Properties and antioxidant activity of probiotic yoghurt flavored with black carrot, pumpkin and strawberry. *Intern. J. Dairy Sci.*, **8**, 48-57.
- AOAC (2000) Official Methods of Analysis, 17th ed. Association of Official Analytical Chemists, Inc. USA.
- **APHA** (2001) American Public Health Association. *Compendium of Methods for the Microbiological Examination of Foods* (4th ed.). Washington D.C.
- Atallah, A.A. (2015) Development of new functional beverages from milk permeate using some probiotic bacteria and fruits pulp. *Egyptian J. Dairy Sci.*, 43, 111-125.
- Bodyfelt, F.W., Tobias, J. and Trout, G.M. (1988) The Sensory Evaluation of Dairy Products. pp. 227-270.Von Nostrand Reinhold. New York.
- Bor, J.Y., Chen, H.Y. and Yen, G.C. (2006) Evaluation of antioxidant activity and inhibitory effect on nitric oxide production of some common vegetables. *Journal of Agricultural and Food Chemistry*, 54,1680-1686.
- Borsheim, E., Tipton, K.D., Wolf, S.E. and Wolfe, R.R. (2003) Essential amino acids and muscle protein recovery from resistance exercise. Am. J. Physoil. Endocrinol. Metab., 283, 648-657.
- Buriti, F.C.A., Freitas, S.C., Egito, A.S. and dos Santos, K.M.O. (2014) Effects of tropical fruit pulps and partially hydrolysed galactomannan from Caesalpinia pulcherrima seeds on the dietary fiber content, probiotic viability, texture and sensory features of goat dairy beverages. *LWT - Food Science and Technology*, **59**,196.
- Caili, F., Haijun, T., Tongyi, C., Yi, L. and Quanhong, L. (2007) Some properties of an acidic protein-bound polysaccharide from the fruit of pumpkin. *Food Chem.*, 100, 944-947.
- **Clarke, G.M. and Kempson, R.E. (1997)** Introduction to the Design and Analysis of *Experiments.* Arnold, a Member of the Holder Headline Group, 1st ed., London, UK.

- Cordenunsi, B.R., Do Nascimento, J.R.O., Genovese, M.I. and Lajolo, F.M. (2002) Influence of cultivar on quality parameters and chemical composition of strawberry fruits grown in Brazil. J. Agric. Food Chem., 50, 2581-2586.
- Elliker, P.R., Anderson, A.W. and Hannesson, G. (1956) An agar medium for lactic acid streptococci and lactobacilli. J. Dairy Sci., 39,1611.
- FAO/WHO, (2002) Guidelines for the evaluation of probiotics in food. Food and Agriculture Organization of the United Nations and World Health Organization Working Group Report. Available from: http://www.fao.org/es/ESN/food/foodandfood_probio_en.stm>.
- Fellers, P.J., Jager, G.De., Pool, M.J., Hill, E.C. and Mittal, P. (1986) Quality of florida-packed retail grapefruit juices as determined by consumer sensory panel and chemical and physical analysis. *J. Food Sci.*, **51**, 417.
- Folkenberg, D.M., Dejmek, P., Skriver, A. and Ipsen, R. (2005) Relation between sensory texture properties and exopolysaccharide distribution in set and in stirred yoghurts produced with different starter cultures. J. Texture Studies, 36,174.
- Geilman, W.G., Schmidt, D., Herfurth-Kennedy, C., Path, J. and Cullor, J. (1992) Production of an electrolyte beverage from milk permeates. *J. Dairy Sci.*, **75**, 2364.
- Gronowska-Senger, A., Smaczny, E. and Dobkowicz, B. (1980) The effect of fiber on utilization of carotene by laboratory rats. *Bromatological Chemia Teksykoloiczna*, 13,129.
- Gupta, R.K. (1998) Handbook of Export Oriented Food Processing Projects. SBP Consultants and Engineers Pvt. Ltd.
- Harvey, T.C. and Catherine, G.C. (1982) Aseptically packaged papaya and guava pulp: Changes in chemical and sensory quality during processing and storage. J. Food Sci., 47, 1164.
- Hashmi, S.I., Kadam, S.S. and Kale, R.V. (2011) Studies on effect of different varieties of date palm paste incorporation on quality characteristics of yoghurt. *Elec. J. Environ., Agric. Food Chem.*, 10, 2371.
- Heenan, C. N., Adams, M. C., Hosken, R. W. and Fleet, G. H. (2004) Survival and sensory acceptability of probiotic microorganisms in a nonfermented frozen vegetarian dessert. *LWT – Food Science and Technology*, 37, 461.
- Hegazi, N.A., AboelEinen, K.A.M. and El-Nawawy, M.A. (2009) Production of low calories highly antioxidant thirst quenching fruit beverage using milk permeate. J. *Biol. Sci.*, 4, 857.
- Hernandez-Ledesma, B., Ramos, M. and Gomez-Ruiz, J.A. (2011) Bioactive components of voine and caprine cheese whey. *Small Ruminant Research*, 101, 196-204.
- **IDF** (1990) Milk and milk products: Enumeration of Yeasts and Moulds (Colony count technique at 25°C). IDF: 94B.

- **IDF** (1991) Milk and milk products: Enumeration of Microorganisms' colony count technique at 30°C. IDF: 100 B.
- Isolauri, E. (2004) The role of probiotics in paediatrics. Current Paediatrics, 14, 104.
- Kalt, W. (2005) Effects of production and processing factors on major fruit and vegetable antioxidants. *J. Food Sci.*, **70**, 12-19.
- Layman, D.K. (2003) The role of leucine in weight loss diets and glucose homeostasis. J. Nutr., 133, 261-267.
- Li, T.S.C. (2008) Vegetables and Fruits: Nutritional and Therapeutic Values. CRC Press, Florida, USA.
- Li, Y., Xu, C., Zhang, Q., Liu, J.Y. and Tan, R.X. (2005) In vitro anti-Helicobacter pylori action of 30 Chinese herbal medicines used to treat ulcer diseases. J. Ethnopharmacol., 98, 329-333.
- Martin, J. H. and Chou, K. M. (1992) Selection of bifidobacteria for use as dietary adjuncts in cultures dairy foods: I-Tolerance to pH of yoghurt. *Cultured Dairy Products Journal*, 21-26.
- Miller, G.D. (2000) Handbook of Dairy Food and Nutrition. 2nd ed. CRC Press LLC. Boca Raton, Florida 33431. P. 24-28.
- Nawirska-Olszanska, A., Biesiada, A., Sokol-Letowska, A. and Kucharska, A.Z. (2011) Content of bioactive compounds and antioxidant capacity of pumpkin puree enriched with Japanese quince, cornelian cherry, strawberry and apples. *Acta Sci. Pol., Technol. Aliment.*, 10, 51-60.
- Prieto, P., Pineda, M. and Aguilar, M. (1999) Spectrophotometric quantitation of antioxidant capacity through the formation of a phosphomolybdenum complex: Specific application to the determination of vitamin E. *Analytical Biochemistry*, 269, 337-341.
- Ryan, M.P., Rea, M.C., Hill, C. and Ross R.P. (1996) An application in cheddar cheese manufacture for a strain of *Lactococcus lactis* producing a novel broad-spectrum bacteriocin, Lacticin 3147. *Appl. Envirn. Microbiol.*, 62, 612.
- Sanmartin, B., Diaz, O., Rodriguez-Turienzo, L. and Cobos, A. (2011) Composition of caprine whey protein concentrates produced by membrane technology after clarification of cheese whey. *Small Ruminant Research*, 105, 186-192.
- Shiri, M.A., Ghasemnezhad, M., Bakhshi, D. and Dadi, M. (2011) Changes in phenolic compounds and antioxidant capacity of fresh-cut table grape (*Vitis vinifera*) cultivar "Shahaneh" as influenced by fruit preparation methods and packagings. *Australian J. Crop Sci.*, 5, 1515-1520.
- Vahedi, N., Tehrani, M.M. and Shahidi, F. (2008) Optimizing of fruit yoghurt formulation and evaluating its quality during storage. Am. Euras. J. Agric. Environ. Sci., 3, 922.

- Walsh, M.K. and Brown, R.J. (2000) Use of amino acid analysis for estimating the individual concentrations of proteins in mixtures. J. Chromatogr. A., 891, 355-360.
- Wolfe, R.R. (2002) Regulation of muscle protein by amino acids. J. Nutr., 132, 3219-3224.
- Wrick, K. (2003) Beverage fortification my how you've changed beverage industry's product development section, 7, 53.
- Zhiyong H., Yuan, B., Maomao, Z., Guanjun, T. and Chen, J. (2015) Effect of simulated processing on the antioxidant capacity and *in vitro* protein digestion of fruit juice-milk beverage model systems. *Food Chemistry*, 457–464.

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إنتاج المشروبات الغذائية الداعمة للحيوية والمدعمة بالمركبات الحيوية النشطة ومضادات الأكسدة في لب اليقطين و الفراولة

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عطالله عبد الرازق عطالله قسم علوم الأغذية – كلية الزراعة – جامعة بنها – ١٣٧٣٦ مشتهر، محافظة القليوبيه – مصر.

في هذه الدراسة ، تم تحضير مشروبات حيوية بواسطة خلط راشح اللبن المتخمر مع لب اليقطين (القرع العسلي) و الفراولة بنسبة خلط (1:1). وتَّم تخمير راشح اللَّبن مستخدماً 2٪ خليط من ثلاث مزارع بادئات بنسب خلط (1:1:1) والتي تتكون من سلالة Lactobacillus delbrueckii subsp bulgaricus DSM 20080 و سلالة Streptococcus thermophilus ASCC 1275 المنتجة للسكريات العديدة و سلالة البكتريا الداعمة للحيوية الـ Bifidobacterium longum DSM 20088 . تم خلط راشح اللبن المتخمر الحلو سواء المعامل حراريا (85°م /51ق) و الغير معامل حراريا بصورة منفصلة مع حجم مساوى له من لب اليقطين أو الفراولة، وتم تعبئة المشروبات المحضرة في زجاجيات معقمة ثم تم تخزينها لمدة 30 يوما على درجة حرارة 4±1م°. تم تقدير التغيرات التى حدثت في المركبات النشطة حيويا و مضادات الأكسده وكذلك الخواص الكميائية ، الميكروبيوجية ، الريولوجية والتقييم الحسى للمشروبات الوظيفية . ووجد زيادة في محتوى المشروبات المحضرة في المواد الصلبة الكلية ومحتوى كل من الدهن ، البروتين ، الرماد ، الالياف و الحموضة بينما انخفض محتواها من الكربو هيدرات الكلية، مضادات الأكسدة، الفينول الكلي ، الفلافونات الكلية ، حمض الاسكوربيك ، الأنثوسيانين ، الكاروتينات و قيم الـ pH خلال فترة التخزين تحت تبريد . وقد وجد أن إضافة كل من لب اليقطين و الفراولة أدى إلى حدوث زيادة كبيرة ولكن بنسب مختلفة في محتواها من الكالسيوم ، الفوسفور ، الصوديوم ، البوتاسيوم ، الماغنسيوم ، الحديد ، النحاس و الزنك للمشروبات المنتجة . ولم يلاحظ وجود أى أعداد من سلالات بكتريا حمض اللاكتيك في المشروبات المنتجة من تسخين راشح اللبن المتخمر وذلك خلال فترة التخزين بالتبريد بينما حدث انخفاض تدريجي في العد Str. thermophilus, Lb. الكلى للبكتريا و بكتريا حمض اللاكتيك و delbrueckii subsp. bulgaricus and Bif. longum في المشروبات المنتجة من راشح اللبن المتخمر غير المسخن خلال فترة التخزين. في حين ظلت أعداد البكتريا الداعمة للحيوية المستخدمة في المشروبات المعدة من راشح اللبن غير المسخن أعلى من العدد المسموح به (log cfug⁻¹) لتحقيق التأثير المفيد والصحى حتّى نهاية فترة التخزين . وقد أوضَحَ التقييمَ الحسى للمشروبات الوظيفية المنتجة ارتفاع درجات القبول الحسي. وقد وجد أن إضافة سلالة البكتريا المنتجة للسكريات العديدة أدت إلى تحسين لزوجة المشروبات الوظيفية المعدة.